

## Structure Determination of Ribosylated Rifampicin and Its Derivative: New Inactivated Metabolites of Rifampicin by Mycobacterial Strains

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(Received for publication April 7, 1995)

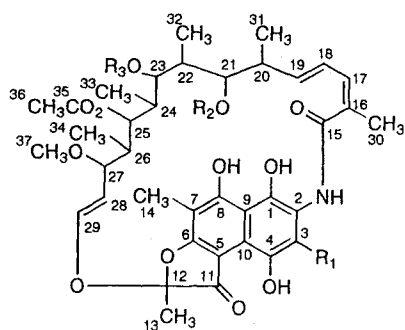
Rifampicin (**1**) was converted into two inactivated products RIP-Ma and RIP-Mb by *Mycobacterium smegmatis* DSM43756. MS, NMR and chromatographic analysis showed the compounds to be 3-formyl-23-[*O*-( $\alpha$ -D-ribofuranosyl)]rifamycin SV (**6**) and 23-[*O*-( $\alpha$ -D-ribofuranosyl)]rifampicin (**7**), respectively.

A semisynthetic antibiotic rifampicin (**1**) is an important chemotherapeutic agent for tuberculosis and leprosy.<sup>1-3</sup> As the inactivated product of rifampicin by *Nocardia* spp., most of which are resistant to rifampicin, we isolated four metabolites RIP-1, -2, -3 and -4, indicating the major resistance mechanism of these organisms is the transformation of **1** to the inactive compounds.<sup>4-6</sup> The inactivation pattern is genus specific<sup>7</sup>: *N. brasiliensis* glycosylate **1** at 23-OH to form

3-formyl-23-[*O*-( $\beta$ -D-glucopyranosyl)]rifamycin SV (RIP-1, **2**) and 23-[*O*-( $\beta$ -D-glucopyranosyl)]rifampicin (RIP-2, **3**), and *N. otitidiscaviarum* phosphorylate **1** at 21-OH to form 21-(*O*-phosphoryl)rifampicin (RIP-3, **4**) and 3-formyl-21-(*O*-phosphoryl)rifamycin SV (RIP-4, **5**) (Fig. 1). *Bacillus* species also inactivate rifampicin by phosphorylation to RIP-3 and RIP-4, or decolorization.<sup>8</sup>

During our studies on rifampicin inactivation by fast

Fig. 1. Structures of rifampicin (**1**), RIP-1 (**2**), RIP-2 (**3**), RIP-3 (**4**), RIP-4 (**5**), RIP-Ma (**6**) and RIP-Mb (**7**).



Rifampicin ( <b>1</b> )	R <sub>1</sub> = CH=N-N <sub>1</sub> (piperazine ring) N-CH <sub>3</sub> R <sub>2</sub> = H, R <sub>3</sub> = H
RIP-1 ( <b>2</b> )	R <sub>1</sub> = CHO, R <sub>2</sub> = H, R <sub>3</sub> = $\beta$ -D-glucose
RIP-2 ( <b>3</b> )	R <sub>1</sub> = CH=N-N <sub>1</sub> (piperazine ring) N-CH <sub>3</sub> R <sub>2</sub> = H, R <sub>3</sub> = $\beta$ -D-glucose
RIP-3 ( <b>4</b> )	R <sub>1</sub> = CH=N-N <sub>1</sub> (piperazine ring) N-CH <sub>3</sub> R <sub>2</sub> = -PO <sub>3</sub> H <sub>2</sub> , R <sub>3</sub> = H
RIP-4 ( <b>5</b> )	R <sub>1</sub> = CHO, R <sub>2</sub> = -PO <sub>3</sub> H <sub>2</sub> , R <sub>3</sub> = H
RIP-Ma ( <b>6</b> )	R <sub>1</sub> = CHO, R <sub>2</sub> = H, R <sub>3</sub> = $\alpha$ -D-ribose
RIP-Mb ( <b>7</b> )	R <sub>1</sub> = CH=N-N <sub>1</sub> (piperazine ring) N-CH <sub>3</sub> R <sub>2</sub> = H, R <sub>3</sub> = $\alpha$ -D-ribose

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growing mycobacterial strains,<sup>9,10</sup> we found that several of them are resistant to rifampicin. From the culture broth of rifampicin resistant *M. smegmatis* DSM43756 grown in the presence of rifampicin, two new inactivated metabolites, RIP-Ma (6) and RIP-Mb (7), were isolated. This paper reports on the structure determination of 6 and 7 (Fig. 1). Biological transformation of rifampicin to RIP-Ma and RIP-Mb, isolation and antimicrobial activities are described elsewhere.<sup>11</sup>

## Experimental

### General

<sup>1</sup>H and <sup>13</sup>C NMR spectra were measured in CD<sub>3</sub>OD on a JEOL ALPHA-500 NMR spectrometer at 500 and 125 MHz, respectively. Chemical shifts of <sup>1</sup>H and <sup>13</sup>C NMR were recorded in  $\delta$  units relative to internal tetramethylsilane ( $\delta=0$ ). FAB-MS and HRFAB-MS were measured on a JEOL JMS-HX110 instrument. HPLC was performed on a Tosoh CCPP-M apparatus.

### Determination of Ribose Configuration

The configuration of ribose in RIP-Mb (7) was determined on a chiral HPLC column after hydrolysis and derivatization. One mg of 7 was hydrolyzed with 2 M-CF<sub>3</sub>COOH (0.2 ml) at 120°C for 1 hour and, after the removal of the solvent with N<sub>2</sub> stream, the reaction product was dried *in vacuo*. The crude ribose was converted to oxime with HONH<sub>2</sub>·HCl (1 mg), CH<sub>3</sub>COONa (2 mg) and methanol (0.2 mg) at 60°C for 1 hour. After removal of the solvent, the product was treated with (CH<sub>3</sub>CO)<sub>2</sub>O (0.2 ml) at 120°C for 1 hour. Removal of the residual reagent followed by dissolving the products in CHCl<sub>3</sub> and washing with H<sub>2</sub>O gave crude aldononitrile acetate (8). After purifying by HPLC (ODS column, mobile phase: 45% CH<sub>3</sub>CN - H<sub>2</sub>O), the ribose derivative 8 was analyzed on a chiral column (Chiralcel OD-H, 4.6 mm i.d. × 250 mm, Daicel Chemical Ind., Ltd., mobile phase: 2% isopropanol - *n*-hexane, 0.5 ml/minute).

## Results and Discussion

### Structure Determination

The structures of RIP-Ma and RIP-Mb were determined by comparison with the spectroscopic data of rifampicin, RIP-1 and RIP-2.<sup>4</sup>

The molecular weights and molecular formulae of RIP-Ma (C<sub>43</sub>H<sub>55</sub>NO<sub>17</sub>: 857) and RIP-Mb (C<sub>48</sub>H<sub>66</sub>N<sub>4</sub>O<sub>16</sub>: 954) were determined by positive and negative FAB-MS and HRFAB-MS (Table 1). These data suggested that RIP-Ma and RIP-Mb are the derivatives of 3-formylrifamycin SV and rifampicin glycosylated with a pentose, respectively.

The structure of RIP-Ma (6) was elucidated by NMR data. <sup>1</sup>H and <sup>13</sup>C NMR chemical shifts were assigned by COSY, C-H COSY and HMBC experiments (Tables 2 and 3). Signals due to a formyl group at  $\delta_H$  10.53 ppm and  $\delta_C$  193.1 ppm were shown in the expense of the signals due to N-methylpiperazine moiety, indicating that 1-amino-4-methylpiperazine has been cleaved off. The presence of ribofuranosyl moiety was indicated by the <sup>1</sup>H and <sup>13</sup>C NMR signals<sup>12~14</sup>:  $\delta_H$  5.24 ppm (H-1'), 3.93 ppm (H-2'), 3.87 ppm (H-3'), 4.01 ppm (H-4') and 3.52 ppm (H-5' × 2): <sup>1</sup>H-<sup>1</sup>H couplings.  $J_{1'-2'}=4.5$  Hz,  $J_{2'-3'}=5.5$  Hz,  $J_{3'-4'}=1.5$  Hz,  $J_{4'-5'}=4.0$  Hz:  $\delta_C$  105.1 ppm (C-1'), 73.2 ppm (C-2'), 71.6 ppm (C-3'), 86.7 ppm (C-4') and 63.4 ppm (C-5'). The glycosylation site was determined by HMBC experiment to correlate H-23 with C-1', and H-1' with C-23 (Fig. 2). Down field shifts of the H-23 signal ( $\Delta\delta_H$  0.14 ppm) and C-23 signal ( $\Delta\delta_C$  9.9 ppm) relative to those of rifampicin were also in accord with the ribosylation of the 23-OH.

The structure of RIP-Mb (7) was similarly determined by NMR data (Tables 2 and 3). <sup>1</sup>H and <sup>13</sup>C NMR spectra of 7 exhibited the signals due to ribosyl group:  $\delta_H$  5.22 ppm (H-1'), 3.93 ppm (H-2'), 3.88 ppm (H-3'), 4.00 ppm (H-4') and 3.51 ppm (H-5'):  $\delta_C$  105.1 ppm (C-1'),

Table 1. Mass spectral data of RIP-Ma (6) and RIP-Mb (7).

	Molecular formula & molecular weight	Positive FAB-MS	Negative FAB-MS	HRFAB-MS
Rifampicin	C <sub>43</sub> H <sub>58</sub> N <sub>4</sub> O <sub>12</sub> 822			
RIP-Ma	C <sub>43</sub> H <sub>55</sub> NO <sub>17</sub> 857	m/z 857 (M) 858 (M+H) 880 (M+Na)	m/z 856 (M-H)	Calcd. for C <sub>43</sub> H <sub>55</sub> NO <sub>17</sub> Na (M+Na) 880.3368 Found 880.3449 (M+Na)
RIP-Mb	C <sub>48</sub> H <sub>66</sub> N <sub>4</sub> O <sub>16</sub> 954	m/z 954 (M) 977 (M+Na) 993 (M+K)	m/z 953 (M-H)	Calcd. for C <sub>48</sub> H <sub>66</sub> N <sub>4</sub> O <sub>16</sub> Na (M+Na) 977.4372 Found 977.4459

Table 2.  $^1\text{H}$  NMR chemical shifts ( $\delta$ , ppm), multiplicity and coupling constants ( $J$ , Hz) of rifampicin (1), RIP-Ma (6) and RIP-Mb (7) in  $\text{CD}_3\text{OD}$ .

Proton	Rifampicin* <sup>1</sup>	RIP-Ma	RIP-Mb
13	1.71(3H, s)	1.70(3H, s)	1.70(3H, s)
14	2.02(3H, s)	2.01(3H, s)	2.02(3H, s)* <sup>4</sup>
17	6.35(1H, br d, 10.5)	6.28(1H, br d, 11.0)	6.36(1H, d, 11.0)
18	7.25(1H, dd, 15.8, 10.5)	7.40(1H, dd, 16.0, 11.0)	7.17(1H, dd, 15.5, 11.0)
19	6.08(1H, dd, 15.8, 7.0)	6.14(1H, dd, 16.0, 7.5)	6.10(1H, dd, 15.5, 7.0)
20	2.31(1H, m)	2.27(1H, ddq, ca. 9.0, 7.0, 7.0)	2.27(1H, m)
21	3.87(1H, dd, 10.0, 1.0)	3.84(1H, dd, 9.5, 1.0)	3.80(1H, brd, 9.0)
22	1.74(1H, m)	1.87(1H, brq, 7.0)	1.80(1H, m)
23	3.08(1H, dd, 10.5, 2.0)	3.22(1H, dd, 9.5, 1.0)	3.21(1H, dd, 9.0, 1.5)
24	1.48(1H, m)	1.62(1H, dq, 9.5, 7.0)	1.65(1H, m)
25	5.16(1H, d, 10.5)	4.96(1H, d, 10.5)	4.97(1H, d, 10.5)
26	1.24(1H, m)	1.15(1H, ddq, 10.5, 1.0, 7.0)	1.17(1H, m)
27	3.38(1H, d, 8.0)	ca. 3.34* <sup>2</sup>	3.30(1H, d, ca. 8.0)* <sup>3</sup>
28	5.07(1H, dd, 12.7, 8.0)	5.08(1H, dd, 12.5, 8.0)	5.11(1H, dd, 12.5, 8.0)
29	6.26(1H, d, 12.7)	6.25(1H, d, 12.5)	6.24(1H, d, 12.5)
30	2.02(3H, s)	1.99(3H, s)	2.01(3H, s)* <sup>4</sup>
31	0.93(3H, d, 7.0)	1.01(3H, d, 7.0)	0.94(3H, d, 7.0)
32	0.99(3H, d, 7.0)	1.03(3H, d, 7.0)	1.01(3H, d, 7.0)
33	0.61(3H, d, 7.0)	0.64(3H, d, 7.0)	0.61(3H, d, 7.0)
34	-0.21(3H, d, 7.0)	-0.12(3H, d, 7.0)	-0.05(3H, d, 7.0)
36	2.02(3H, s)	2.01(3H, s)	2.02(3H, s)* <sup>4</sup>
37	3.00(3H, s)	3.00(3H, s)	3.00(3H, s)
N-CH <sub>3</sub>	2.78(3H, s)	----	2.60(3H, s)
PhCH=N-N	8.32(1H, s)	----	8.24(1H, s)
PhCH=O	----	10.53(1H, s)	----
CH <sub>2</sub> N	3.30(4H, br m)* <sup>3</sup>	----	3.26(4H, brm)
	3.18(4H, br m)	----	2.95(4H, brm)
1'		5.24(1H, d, 4.5)	5.22(1H, d, 4.5)
2'		3.93(1H, dd, 5.5, 4.5)	3.93(1H, dd, 5.5, 4.5)
3'		3.87(1H, dd, 5.5, 1.5)	3.88(1H, dd, 5.5, 2.0)
4'		4.01(1H, ddd, ca. 4.0, 4.0, 1.5)	4.00(1H, m)
5'		3.52(2H, d, 4.0)	3.51(2H, d, 4.0)

\*<sup>1</sup> N. MORISAKI *et al.*, J. Antibiotics, 46: 1605~1610, 1993.\*<sup>2</sup> Overlapping with other signals.\*<sup>3</sup> Obscured by other resonances.\*<sup>4</sup> Interchangeable.

73.2 ppm (C-2'), 71.7 ppm (C-3'), 86.7 ppm (C-4') and 63.5 ppm (C-5'), that were absent in the spectra of rifampicin (1). Down field shifts of H-23 ( $\Delta\delta_{\text{H}}$  0.13 ppm) and C-23 signal ( $\Delta\delta_{\text{C}}$  9.7 ppm) of 7 relative to those of rifampicin indicated the ribosylation of the OH group on C-23.

The absolute configuration of the ribose was determined on a chiral HPLC column (Scheme 1). The ribose

obtained by hydrolysis of RIP-Mb (7) was converted to the aldononitrile acetate 8,<sup>15)</sup> and 8 was analyzed on Chiralcel OD-H column.<sup>16)</sup> Retention time of the derivatives 8 from the ribose of RIP-Mb, L-ribose and D-ribose were 60.0 minutes, 58.0 minutes and 59.6 minutes, respectively. Accordingly, the absolute configuration of ribose in RIP-Mb was assigned to be D.

The configuration at anomeric carbon of D-ribose in

Table 3.  $^{13}\text{C}$  NMR chemical shifts ( $\delta$  ppm) of rifampicin (1), RIP-Ma (6) and RIP-Mb (7) in  $\text{CD}_3\text{OD}$ .

Carbon	Rifampicin* <sup>1</sup>	RIP-Ma	RIP-Mb	Carbon	Rifampicin	RIP-Ma	RIP-Mb
1-10	184.0	185.2	184.8	25	75.6	75.4	75.3
	175.8	174.8	174.8	26	41.7	42.1	42.3
	149.3	152.5	149.2	27	78.5	78.4	78.5
	147.9	149.1	147.5	28	120.1* <sup>3</sup>	120.0	120.0
	119.7* <sup>3</sup>	120.7	119.5	29	144.7	144.7	144.7
	118.2	120.2	118.4	30	20.8	20.3	20.7
	116.1	119.4	116.5	31	18.2	18.1	18.3
	116.0	115.2	116.0	32	11.0	11.6	11.7
	105.0	105.7	104.6	33	9.4	10.3	10.2
	101.9	101.6	101.5	34	9.7	9.5	9.7
11	189.0	188.9	188.2	35	172.4	172.5	172.5
12	110.6	110.4	110.4	36	20.8	21.0	21.0
13	22.4	22.2	22.3	37	56.7	56.7	56.7
14	7.5	7.4	7.5	NCH <sub>3</sub>	43.7	---	44.6
15	171.1	171.8	170.6	PhCH=N-N	138.8	---	137.7
16	133.1	133.4	133.3	PhCH=O	---	193.1	---
17	134.8	134.0	134.8	CH <sub>2</sub> N	53.2	---	53.8
18	129.0	129.7	129.0		49.5* <sup>2</sup>	---	50.1
19	140.7	140.0	141.1	1'		105.1	105.1
20	39.1	38.9	39.4	2'		73.2	73.2
21	75.2	76.5	75.6	3'		71.6	71.7
22	34.4	34.8	35.0	4'		86.7	86.7
23	78.2	88.1	87.9	5'		63.4	63.5
24	39.4	39.6	39.6				

\*<sup>1</sup> N. MORISAKI *et al.*, J. Antibiotics, 46: 1605~1610, 1993.\*<sup>2</sup> Obscured by other resonances.\*<sup>3</sup> Interchangeable.

Scheme 1. Derivatization of ribose in RIP-Mb (7) for HPLC analysis.

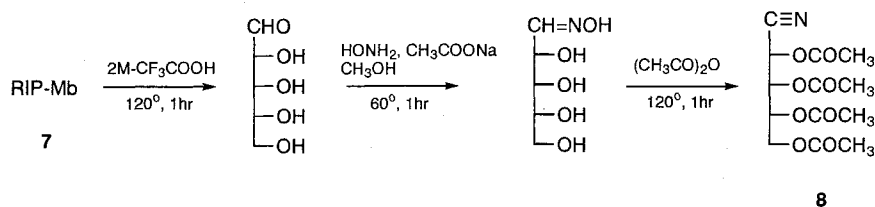
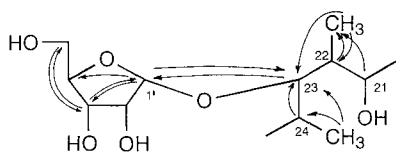


Fig. 2. Partial structure of RIP-Ma.

Arrows indicate  $^1\text{H}$ - $^{13}\text{C}$  correlation by HMBC experiment.

RIP-Ma and RIP-Mb was determined to be  $\alpha$  from the reported  $^{13}\text{C}$  NMR chemical shift data of methyl  $\alpha$ - and  $\beta$ -furanosides of pentoses.<sup>12~14)</sup> Thus, the structures of RIP-Ma (6) and RIP-Mb (7) were determined to be 3-formyl-23-[*O*-( $\alpha$ -D-ribofuranosyl)]rifamycin SV and 23-[*O*-( $\alpha$ -D-ribofuranosyl)]rifampicin, respectively. During the incubation period, the ratio of RIP-Ma and RIP-Mb in the culture medium was almost constant, and RIP-Mb did not decompose to RIP-Ma in the isolation

and purification procedure. Therefore, the imide group is considered to be biologically hydrolyzed.

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